

Composition and modification of the lung microbiome in patients with idiopathic pulmonary fibrosis

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ABSTRACT



Lungs have long been considered sterile, but recent research has shown that a large number of microbiological organisms exist in the lungs of healthy subjects (including bacteria, fungi, and viruses), collectively known as the microbiome. It undergoes changes in patients with respiratory pathology. Studies in idiopathic pulmonary fibrosis show that a large number of bacteria or the abundant presence of potentially pathogenic bacteria can cause disease progression and exacerbation, and can implicitly increase mortality. There seems to be a quantitative balance, a well-established proportion between the components of this microbiome, which is disturbed during a disease and can reach a "state of pulmonary dysbiosis". Evidence suggests that the microbiome may be used as a prognostic biomarker and may also explain the pathogenesis of interstitial fibrosis.

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Introduction

The microbiome consists of microorganisms (and their products) that are physiologically found in the human body [1]. The human microbiota is made up of trillions of symbiotic microbial cells, mainly bacteria in the intestine; in addition to this, viruses (including bacteriophages), fungi, and archaea (archaeobacteria, part of the oral, intestinal and vaginal microbiome) are also identified [1,2]. Projects on the microbiome, from all over the world, have been launched in order to understand the roles that these germs play in the appearance and evolution of different pathologies, as well as their impact on human health. The human body and microorganisms have coexisted for millions of years, so the immune system of the human body and the microbiome are closely linked [1,3]. Apparently, each compartment of the human body has a unique

microbiome made up of specific microorganisms, and consequently the respiratory tree has its own microbiome [1].

The purpose of the current review is to analyze idiopathic pulmonary fibrosis, a disease with fatal impact, from the point of view of the pulmonary microbiome and the benefits it may bring to our knowledge in the evolution of the disease.

Discussions

The experimental part

Studies that aimed to determine the composition of the microbiome in the healthy man, in the patient with diagnosed idiopathic pulmonary fibrosis, and the modification of the microbiome in the context of the evolution of the disease and exacerbations (where the patients' condition allowed) were analyzed.

Most of the studies that investigated the human microbiome insisted on the intestinal one, considering the wide range of bacteria found at this level. Also, in the oropharynx, skin, vagina, and nasal cavity, the microbiome was easier studied in comparison with the lung, due not only to greater accessibility, but also to the large number of present microorganisms [4].

The microbiome controversy is about the physiological colonization of the respiratory tree. Lungs have long been considered sterile because standard microbiological culture techniques consistently give negative results [5], only 1% of bacteria can be grown in normal laboratories [6]. In the last decade, however, the use of molecular techniques has shown that this theory is incorrect and that a large number of microbiological organisms (including bacteria, fungi, and viruses), collectively known as the microbiome, coexist in the lungs not only of healthy subjects, but also in patients with respiratory diseases [5].

The challenges for describing the respiratory microbiome are increasing and take into account the difficulty obtaining samples from the lower respiratory tract as well as their contamination in the upper respiratory tract [1].

The respiratory mucosa is host to numerous microorganisms. The lung microbiome evaluation studies were based on the analysis of bronchoalveolar lavage obtained by bronchoscopy [1]. This technique is widely used to study and highlight the pathogenetic mechanisms and inflammatory response through the influx of pro-inflammatory cells [7]. Although sputum examination is a less invasive method compared to bronchoscopy, it presents an increased risk of contamination when passing through the oropharyngeal cavity [1].

The most representative bacteria in the respiratory tract have so far proved to be: Bacteroidetes (including *Prevotella* sp.), Firmicutes (including *Streptococcus* sp. and *Veillonella* sp.) and, in lower quantities, Proteobacteria and Actinobacteria [8–10]. It is worth mentioning that despite the difference in pH, temperature, and oxygen concentration, the microbiome in healthy subjects is relatively constant.

The composition of the microbiome is influenced by mucociliary clearance and host defense mechanisms, local growth factors contributing very little to the composition of the microbiome in healthy lungs [9]. Smoking can directly affect the composition of the microbiome, which can lead to changes in the structure of microorganisms [11].

Various types of bacteria promote inflammation in the respiratory tract. Although they are less common (*Haemophilus influenzae*, *Moraxella catarrhalis*, *Streptococcus pneumoniae*, *Haemophilus parainfluenzae*, *Staphylococcus aureus*, and *Pseudomonas aeruginosa*), their persistence in the respiratory tract favors chronic

respiratory diseases and chronic inflammation [12–14]. These microorganisms become particular in the case of certain pathologies, and the overlap and agglomeration of bacterial species decrease the physiological species richness of the pulmonary microbiota and are associated with the evolution of some diseases [15].

All of these bacterial species carry a unique imprint, a distinct complement of genomic DNA. Since it is impossible to completely sequence each genome in each cell, microbial ecology has defined a series of molecular markers that express a DNA sequence which identifies that particular genome [16].

Several markers have been evaluated over time, including ribosomal protein subunits, growth factors, and RNA polymerase subunits. The most present and historically significant is the small or 16S ribosomal RNA subunit gene, commonly referred to as 16s rRNA [16,17]. This subunit is present in the genome of all bacteria, making it easy to identify bacteriological sequences and is the target site for amplification or sequencing [18]. The rRNA-16s sequencing identifies bacterial DNA in 97% of the cases that have benefited from bronchial lavage, compared to the conventional culture technique, which detects bacteria in 39.1% of the lavage sample [8].

Pulmonary microbiome and idiopathic pulmonary fibrosis

Interstitial lung disease (ILD) is a heterogeneous group of conditions with similar clinical, imaging, and histopathological features. While some are associated with immune impairment, others are allergic or even idiopathic. The pathogenesis of these diseases is not fully elucidated; the exaggeration of the host's immune response, which acts as a continuous stimulant to the alveolar epithelium, may be a triggering mechanism of the disease [19]. The pathogenesis of pulmonary fibrosis is complex and may be specific for different agents, with the destruction of pulmonary architecture [20].

Few studies have been carried out on the microbiome in interstitial lung disease. However, more recent studies suggest the importance and connection of the lung microbiome with respiratory pathologies, including interstitial pulmonary fibrosis (IPF) [1]. Recent studies have described the microbiome from stable idiopathic pulmonary fibrosis and correlated the microbial composition and bacterial load with the evolution of pulmonary fibrosis [21]. The lung microbiome was evaluated in patients with IPF in the (COMET) -IPF (Correlating Outcomes with biochemical Markers to Estimate Time-progression) study, and the most prevalent identified bacteria were *Prevotella*, *Veillonella* and *Cronobacter* species (spp.) [22].

As research into the composition of the microbiome in IPF is in its early stages, correlations with intensively studied pathologies (such as COPD) and with healthy

subjects were analyzed through PCR studies from bronchoalveolar lavage. Patients with IPF had a significantly higher bacterial load compared to the COPD group and the healthy group. The most abundant species present in these groups were *Streptococcus*, *Prevotella*, and *Veillonella* spp. and patients with IPF showed lower bacterial diversity compared to the control group, harboring potentially pathogenic bacteria such as *Haemophilus*, *Neisseria* and *Streptococcus* spp. [23]. Among patients with IPF, high bacterial load was associated with a significant decrease in survival time compared to those with lower bacterial load, an effect that is independent of age and tobacco use [23-25].

Interestingly, patients with IPF who have the MUC5B minor allele genotype, previously at increased risk of developing IPF and playing an important role in mucociliary clearance [26,27], had significantly lower bacterial burden compared with patients with IPFs that do not have this genotype [23].

Different studies have evaluated gene expression in blood samples and polymorphisms of MUC5B genes in patients with IPF, compared to healthy control groups. Overexpression of a gene in the IPF group was associated with death and disease progression, with a high level of neutrophils in both peripheral blood and bronchoalveolar lavage, a high bacterial load in bronchoalveolar lavage, and a relatively low amount of *Neisseria* sp [28].

After adjusting the parameters (age, gender, smoking status, presence of gastroesophageal reflux, baseline lung function, and 6-minute walk test), the presence of *Streptococcus* or *Staphylococcus* over a certain limit was associated with a significant clinical reduction in survival. This finding shows that potentially pathogenic microorganisms (*Streptococcus* and *Staphylococcus* sp.) are associated with an increased risk of disease progression in humans, coinciding with exacerbation of pulmonary fibrosis in 2 specimens of mice infected with *Streptococcus pneumoniae* [22].

We must not forget that there are a number of limitations in the evaluation of the lung microbiome in the context of an exacerbation. The worsening of the respiratory condition, with significant desaturations can prevent the obtaining of bronchoalveolar lavage. Also, the administration of the antibiotic before the harvest influences the obtained material.

An acute exacerbation of idiopathic pulmonary fibrosis (AE-IPF) has been defined as an acute clinical worsening of dyspnea, which develops in less than 1 month without an alternative etiology.

While AE-IPF is increasingly recognized and perceived as a severe event with high mortality, there is only a limited amount of clinical data on exacerbation of other interstitial lung pathologies [29]. Precisely from this more accurate identification of IPF, the pulmonary microbiome was

further studied in the context of exacerbation of IPF, which does not yet have a definite conclusion. Some studies incriminate *Staphylococcus* and *Streptococcus* in aggravating the underlying disease [19].

Molecular techniques have identified several common pathogenic viruses that appear in exacerbation, including respiratory syncytial virus and cytomegalovirus [30]. Described as molecular techniques applied to sputum cultures of patients with acute exacerbation, 89% of gram-negative bacteria with pathogenic potential were found: *Klebsiella pneumoniae* (26%), *Mycobacterium tuberculosis* (21%), and *Acinetobacter baumannii* (10%) were the dominant sputum samples [31].

PF progression includes exacerbations like other respiratory pathologies. Acute exacerbations have been associated with poor prognosis. Patients who died by exacerbation had a higher percentage of neutrophils in the bronchoalveolar fluid and a lower number of lymphocytes in the same sample, compared to those who survived an exacerbation [32]. The correlation of the neutrophils with the change in the composition of the lung microbiome could be a key for future studies.

Also, the role of acute phase PCR protein has been studied, which has been shown to be an independent predictor of survival, and a viral or bacterial infection and/or inflammation may be one of the pathogenic mechanisms of exacerbation [32,33]. Patients with IPF who experienced an exacerbation were found to have four times greater bacterial load compared to patients with stable IPF from the control group, who met the same criteria for age, gender, smoking status, and lung volume.

Exacerbated patients had a higher colony of Proteobacteria sp and potentially pathogens such as *Campylobacter* and *Stenotrophomonas* spp. [34]; *Campylobacter* is known to be a pathogen that originates from the gastrointestinal tract. This finding supports the idea that gastroesophageal reflux would contribute to exacerbations in IPF [35,36].

Evaluation of the microbiome from the sputum of patients with idiopathic pulmonary fibrosis showed an increase in bacterial load with *Campylobacter* sp. and *Stenotrophomonas* sp., together with a significant decrease of *Veillonella* sp. [34]

The contribution of viral infections, including viral hepatitis C, transfused viruses, and herpes virus has been intensively studied, but with conflicting results regarding their importance in the pathogenesis of the disease [37]. Herpetic viruses (including Epstein-Barr virus, cytomegalovirus, herpes simplex virus, and human herpes virus-7 and -8) have been identified in a large proportion in the lung tissue of IPF patients, compared to the control group [38] and similar viruses have developed fibrosis in experimental animals [37]. These data are influenced by the immunosuppressive treatment given to patients with

IPF, but may suggest that viruses are co-factors in the progression of pulmonary fibrosis [21].

Lowering of respiratory infections is a benefit on overall mortality in patients with IPF who have received antibiotic therapy with cotrimoxazole [39]. The standard treatment of idiopathic pulmonary fibrosis with the main antifibrotic agents (Pirfenidone and Nintedanib) has as a secondary effect the modification of the intestinal motility [39], through the transit modifications that it favors (diarrhea and constipation).

Modification of the intestinal microbiome favors respiratory infections [40], with all the implications described above, practically proving that in patients with IPF, they are very difficult to control. In this context, an ideal therapeutic agent should combine the antimicrobial, antiviral, and antifibrotic effects [41,42], although it remains to be discussed whether alternative therapies would have antiviral, antimicrobial, anti-inflammatory and antifibrotic effects [43,44]

Herbal remedies such as Black Elder (*Sambucus nigra* L.) have been identified so far which associates antiviral (herpes simplex virus type 1, influenza virus) and antimicrobial (*Streptococcus pneumoniae*, *Haemophilus influenza*, *Streptococcus pyogenes*, group C and G *Streptococci*, *Branhamella catarrhalis*, and *Haemophilus influenza*) [45] effects, with demonstrated myocardial antifibrotic effects [46] and renal tissue [47].

Analyzing the previously described aspects, we find that the progression of idiopathic pulmonary fibrosis is correlated with the modification of the respiratory microbiome. There is also some evidence to suggest that viral infection may be responsible for a proportion of acute exacerbations of IPF. The role of the bacteria in the pathogenesis of IPF is less clear. Studies of other respiratory diseases suggest that changes in the lung microbiome are associated with the disease and that these changes influence the behavior of the disease [37].

Conclusions

Interest in the lung microbiome has grown steadily over the past decade, proving through various studies that respiratory microbiota microorganisms play an important role in maintaining the health-disease balance.

Respiratory tract dysbiosis causes an irregular immune response, which in turn can promote host susceptibility to infections, with the deepening microbial imbalance.

In the evolution of chronic lung disease, the microbiome could help select appropriate, targeted, and more personalized antibiotics during the disease, especially in exacerbations of idiopathic pulmonary fibrosis.

In this context, more advanced metagenomic analyses are needed to elucidate the functional role of individual genes and bacterial communities in the progression of idiopathic pulmonary fibrosis.

Conflict of interest disclosure

There are no known conflicts of interest in the publication of this article. The manuscript was read and approved by all authors.

Compliance with ethical standards

Any aspect of the work covered in this manuscript has been conducted with the ethical approval of all relevant bodies and that such approvals are acknowledged within the manuscript.

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All authors have equal rights as the first author of this paper.

References

1. Gill SR, Pop M, DeBoy RT, Eckburg PB, Turnbaugh PJ, Samuel BS, et al. Metagenomic analysis of the human distal gut microbiome. *Science*. 2006; 312(5778):1355–9. doi:10.1126/science.1124234
2. Dridi B, Raoult D, Drancourt M. Archaea as emerging organisms in complex human microbiomes. *Anaerobe*. 2011;17(2):56–63. doi:10.1016/j.anaerobe.2011.03.001
3. Bäckhed F, Ley RE, Sonnenburg JL, Peterson DA, Gordon JI. Host-bacterial mutualism in the human intestine. *Science*. 2005; 307(5717): 1915–1920. doi:10.1126/science.1104816
4. Mitchell AB, Glanville AR. The Human Respiratory Microbiome: Implications and Impact. *Semin Respir Crit Care Med*. 2018;39(2):199–212. doi:10.1055/s-0037-1617441
5. Faner R, Sibila O, Agustí A, et al. The microbiome in respiratory medicine: current challenges and future perspectives. *Eur Respir J*. 2017;49(4):1602086. Published 2017 Apr 12. doi:10.1183/13993003.02086-2016
6. Hilty M, Burke C, Pedro H, et al. Disordered microbial communities in asthmatic airways. *PLoS One*. 2010;5(1):e8578.
7. Domokos-Hancu B, Man MA, Trofor A, et al. Biochemistry in Assessing the Inflammatory Response of the Respiratory System Due to Experimental Exposure to Glass Fibres. *Mater Plast*. 2019; 56(1): 285–290.
8. Dickson RP, Erb-Downward JR, Prescott HC, Martinez FJ, Curtis JL, Lama VN, et al. Analysis of culture-dependent versus culture-independent techniques for identification of bacteria in clinically obtained bronchoalveolar lavage fluid. *J Clin Microbiol*. 2014;52(10):3605–13.
9. Dickson RP, Erb-Downward JR, Freeman CM, McCloskey L, Beck JM, Huffnagle GB, et al. Spatial

- variation in the healthy human lung microbiome and the adapted island model of lung biogeography. *Ann Am Thorac Soc*. 2015;12(6):821–30.
10. Erb-Downward JR, Thompson DL, Han MK, et al. Analysis of the lung microbiome in the "healthy" smoker and in COPD. *PLoS One*. 2011;6(2):e16384.
 11. Tong X, Su F, Xu X, Xu H, Yang T, Xu Q, Dai H, Huang K, Zou L, Zhang W, Pei S, Xiao F, Li Y, Wang C. Alterations to the Lung Microbiome in Idiopathic Pulmonary Fibrosis Patients. *Front Cell Infect Microbiol*. 2019;9:149.
doi: 10.3389/fcimb.2019.00149
 12. Kovaleva OV, Romashin D, Zborovskaya IB, Davydov MM, Shogenov MS, Gratchev A. Human Lung Microbiome on the Way to Cancer. *J Immunol Res*. 2019;2019:1394191. doi:10.1155/2019/1394191
 13. Sethi S, Murphy TF. Bacterial infection in chronic obstructive pulmonary disease in 2000: a state-of-the-art review. *Clin Microbiol Rev*. 2001;14(2):336–363.
doi:10.1128/CMR.14.2.336-363.2001
 14. Zălar DM, Pop C, Buzdugan E, Todea D MC. The Atherosclerosis Inflammation Relationship. A Pathophysiol Approach. *Farmacia*. 2019; 67(6): 941–947. doi: 10.31925/farmacia.2019.6.2
 15. Mathieu E, Escribano-Vazquez U, Descamps D, et al. Paradigms of Lung Microbiota Functions in Health and Disease, Particularly, in Asthma. *Front Physiol*. 2018;9:1168. doi:10.3389/fphys.2018.01168
 16. Morgan XC, Huttenhower C. Chapter 12: Human microbiome analysis. *PLoS Comput Biol*. 2012; 8(12):e1002808.
 17. Tringe SG, Hugenholtz P. A renaissance for the pioneering 16S rRNA gene. *Curr Opin Microbiol*. 2008;11(5):442–6.
 18. Clarridge JE 3rd. Impact of 16S rRNA gene sequence analysis for identification of bacteria on clinical microbiology and infectious diseases. *Clin Microbiol Rev*. 2004;17(4):840–862. doi:10.1128/CMR.17.4.840–862.2004
 19. Costa AN, Costa FMD, Campos SV, Salles RK, Athanazio RA. The pulmonary microbiome: challenges of a new paradigm. *J Bras Pneumol*. 2018;44(5):424–432. doi:10.1590/S1806-37562017000000209
 20. Morris A, Beck JM, Schloss PD, Campbell TB, Crothers K, Curtis JL, et al. Comparison of the respiratory microbiome in healthy nonsmokers and smokers. *Am J Respir Crit Care Med*. 2013; 187(10):1067–75.
 21. Salisbury ML, Han MK, Dickson RP, Molyneaux PL. Microbiome in interstitial lung disease: from pathogenesis to treatment target. *Curr Opin Pulm Med*. 2017;23(5):404–410.
doi:10.1097/MCP.0000000000000399
 22. Han MLK, Zhou Y, Murray S, Tayob N, Noth I, Lama VN, et al. Lung microbiome and disease progression in idiopathic pulmonary fibrosis: An analysis of the COMET study. *Lancet Respir Med*. 2014;2(7):548–56.
 23. Molyneaux PL, Cox MJ, Willis-Owen SAG, Mallia P, Russell KE, Russell AM, et al. The role of bacteria in the pathogenesis and progression of idiopathic pulmonary fibrosis. *Am J Respir Crit Care Med*. 2014;190(8):906–13.
 24. Budin CE, Marginean C, Bordea IR, Enache LS, Enache EL, et al. The Influence of Smoking on Nicotine Exposure Biomarkers and Inflammatory Profile Among Foster Care Teenagers, Romania. *REV. CHIM. (Bucharest)*. 2018; 69(12):3659–63.
 25. Budin CE, Alexescu TG, Bordea IR, Gherghinescu MC, Aluas M, et al. Nicotine Addiction Objective in Educational Programs for Smoking Prevention in Young People. *REV. CHIM. (Bucharest)*. 2019; 70(6): 2168– 72.
 26. Peljto AL, Zhang Y, Fingerlin TE, Shwu-Fan M, Garcia JGN, Richards TJ, et al. Association between the MUC5B promoter polymorphism and survival in patients with idiopathic pulmonary fibrosis. *JAMA*. 2013;309(21):2232–9.
 27. Zhang Y, Noth I, Garcia JG, Kaminski N. A variant in the promoter of MUC5B and idiopathic pulmonary fibrosis. *N Engl J Med*. 2011;364(16):1576–1577.
doi:10.1056/NEJMc1013504
 28. Molyneaux PL, Willis-Owen SAG, Cox MJ, James P, Cowman S, Loebinger M, et al. Host-microbial interactions in idiopathic pulmonary fibrosis. *Am J Respir Crit Care Med*. 2017;195(12):1640–50.
 29. Leuschner G, Behr J. Acute Exacerbation in Interstitial Lung Disease. *Front Med (Lausanne)*. 2017;4:176.
doi:10.3389/fmed.2017.00176
 30. Ushiki A, Yamazaki Y, Hama M, Yasuo M, Hanaoka M, Kubo K. Viral infections in patients with an acute exacerbation of idiopathic interstitial pneumonia. *Respir Investig*. 2014;52(1):65–70.
 31. Weng D, Chen XQ, Qiu H, et al. The Role of Infection in Acute Exacerbation of Idiopathic Pulmonary Fibrosis. *Mediators Inflamm*. 2019;2019:5160694.
doi:10.1155/2019/5160694.
 32. Song JW, Hong SB, Lim CM, Koh Y, Kim DS. Acute exacerbation of idiopathic pulmonary fibrosis: Incidence, risk factors and outcome. *Eur Respir J*. 2011;37(2):356–63.
 33. Alexescu TG, Bordea IR, Cozma A, Rajnoveanu R, Buzoianu AD, Nemes RM, Tudorache SI, Boca BM, Todea DA. Metabolic Profile and the Risk of Early Atherosclerosis in Patients with Obesity and Overweight. *REV. CHIM. (Bucharest)* 2019;70(10): 3627–33.

34. Molyneaux PL, Cox MJ, Wells AU, Kim HC, Ji W, Cookson WOC, et al. Changes in the respiratory microbiome during acute exacerbations of idiopathic pulmonary fibrosis. *Respir Res*. 2017;18(1):29.
35. Raghu G, Collard HR, Egan JJ, Martinez FJ, Behr J, Brown KK, et al. An Official ATS/ERS/JRS/ALAT Statement: Idiopathic pulmonary fibrosis: Evidence-based guidelines for diagnosis and management. *Am J Respir Crit Care Med*. 2011;183(6):788–824.
36. Lee JS, Ryu JH, Elicker BM, Lydell CP, Jones KD, Wolters PJ, et al. Gastroesophageal reflux therapy is associated with longer survival in patients with idiopathic pulmonary fibrosis. *Am J Respir Crit Care Med*. 2011;184(12):1390–4.
37. Molyneaux PL, Maher TM. The role of infection in the pathogenesis of idiopathic pulmonary fibrosis. *Eur Respir Rev*. 2013;22(129):376–381. doi:10.1183/09059180.00000713
38. Tang YW, Johnson JE, Browning PJ, Cruz-Gervis RA, Davis A, Graham BS, et al. Herpesvirus DNA is consistently detected in lungs of patients with idiopathic pulmonary fibrosis. *J Clin Microbiol*. 2003;41(6):2633–40.
39. Rozenberg D, Sitzler N, Porter S, et al. Idiopathic Pulmonary Fibrosis: A Review of Disease, Pharmacological, and Nonpharmacological Strategies With a Focus on Symptoms, Function, and Health-Related Quality of Life. *J Pain Symptom Manage*. 2019; S0885-3924: 31078-4.
40. McAleer JP, Kolls JK. Contributions of the intestinal microbiome in lung immunity. *Eur J Immunol*. 2018;48(1):39–49. doi:10.1002/eji.201646721
41. Gorkiewicz G, Thallinger GG, Trajanoski S, et al. Alterations in the colonic microbiota in response to osmotic diarrhea. *PLoS One*. 2013;8(2):e55817.
42. Ardeleanu V, Francu L, Georgescu C. Neoangiogenesis. Assessment in Esophageal Adenocarcinomas. *Indian J Surg*. 2015;77(Suppl 3):971–976. doi:10.1007/s12262-014-1091-9
43. Maierian A, Ciumarnean L, Alexescu TG, Domokos B, Rajnoveanu R, et al. Complementary therapeutic approaches in asthma. *Balneo Res Journal*. 2019;10(3):204–212.
44. Alexescu TG, Tarmure S, Negrean V, Cosnarovici M, Ruta VM, et al. Nanoparticles in the treatment of chronic lung diseases. *J Mind Med Sci*. 2019; 6(2): 224–231. doi: 10.22543/7674.62.P224231
45. Porter RS, Bode RF. A Review of the Antiviral Properties of Black Elder (*Sambucus nigra* L.). *Phytotherapy Research*. 2017; 31:533–54.
46. Ciocoiu M, Badescu M, Badulescu O, Badescu L. The beneficial effects on blood pressure, dyslipidemia and oxidative stress of *Sambucus nigra* extract associated with renin inhibitors. *Pharm Biol*. 2016;54(12):3063–7.
47. Ungur R, Buzatu R, Lacatus R, Purdoi RC, Petrut G, et al. Evaluation of the Nephroprotective Effect of *Sambucus Nigra* Total Extract in a Rat Experimental Model of Gentamicine Nephrotoxicity. *Rev. Chim*. 2019; 70(6):1971–1974.