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THE GENUS PHRAGMATOBIA IN NORTH AMERICA, WITH THE
DESCRIPTION OF A NEW SPECIES (LEPIDOPTERA: ARCTIIDAE)

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This paper, based on the examination of 1,879 specimens,
serves to resolve the taxonomic problems involving the three North
American species of Phragmatobia. The genus Phragmatobia, the ruby
tiger moths, has had a checkered history since it was described by
Stephens in 1829 (type, by monotypy, Noctua fuliginosa Linnaeus, 1758).
Although many species have been described in or transferred to this
genus, in both the Old and New Worlds, most of them have been re-
moved to other genera. By 1902 Dyar recognized only two North Ameri-
can species, a status since then unchanged (McDunnough, 1938;
Forbes, 1960). Despite the recent stability of the names, there has
been much confusion as to which names to apply to particular speci-
mens. This problem is resolved below, with the description of a third
North American species, long confused with the two named species.

North American Phragmatobia are rosy, stout, hairy, medium-
sized moths, restricted to the Hudsonian, Canadian, Transition, and
northern Upper Austral Zones. As with many arctiids, there is con-
siderable variation in color, especially in the hindwings. Females are
generally darker than males, and are occasionally difficult to identify
with certainty. The males are readily attracted to lights, especially
near-ultraviolet ("black") lights, early in the evening, but the females
are only rarely collected this way. Of 134 specimens of P. fuliginosa
collected in Maine by Dirks (1937), only 7, or 5.2%, were females.
This disparity is even greater for P. assimilans in Michigan, where no
females have ever been collected. Ferguson (pers. comm.) similarly
reports that he does not recall collecting a single female P. assimilans
in Nova Scotia, though he has taken about 100 males. Many of the fe-
males which we examined in the course of this study were reared speci-
mens.

The highly variable "bear" larvae of all three species have been
briefly described, but never adequately illustrated. They usually feed
on a variety of low herbaceous plants, but all can be reared on Taraxa-
cum (dandelion), Plantago (plantain), or Rumex (dock).

Phragmatobia is homogeneous and well-defined in North Ameri-
ca, but the status of the many Palearctic species, forms, varieties,
and aberrations ascribed to this genus is far from clear. The constant
and specific differences we have found in the male genitalia may lead to
a re-evaluation of the Palearctic representatives.
Several Neotropical moths have been described in or transferred to *Phragmatobia*, but we have made no attempt to evaluate their taxonomic status. These include *P. fervida* (Walker, 1855); *P. modesta* Maassen, 1890; *P. nudar* Dyar, 1907; *P. rubricosta* Dognin, 1889; and *Phragmatobia* [sic] viridis Druce, 1903. We have made no attempt to apply to our North American species the names of "forms" and "aberrations" proposed for Palearctic species. These names include "borea is" Staudinger, "fervida" Staudinger (necc. Walker), "subnigra" Millière, and "pulverulenta" Alpheraky, all of which were catalogued under *P. fuliginosa* by Dyar (1902). A careful check has shown that none of these names applies to the new species described here.

The generic characters have been fairly well enumerated by Forbes (1960), although several characters deserve special mention. The male genitalia (Figs. 1-9, 13-15) follow a simple plan (terminology after Klots, 1956): the uncus, tegumen, vinculum, valvae, aedeagus, juxta, and saccus are well developed and relatively unmodified. Most of these structures exhibit specific differences among the three species. The vesica is ornamented with numerous, stout, thorn-like cornuti, while the distal end of the aedeagus is adorned with carinae of taxonomic importance (Figs. 7-9). The valvae (Figs. 13-15) are long and narrow, with a single spade-shaped projection on the mesal face, or with two lobes, one on the dorsal and one on the ventral margin. The shape of these projections is somewhat variable, but their position and number are constant.

The female genitalia (Figs. 20-25) are fairly uniform, and no striking differences between species have been found, although a few minor differences will be pointed out later. The ductus bursae is strongly sclerotized, dorso-ventrally compressed, and almost as wide as the abdomen. The cervix bursae is large, firm, and somewhat convoluted. The corpus bursae is large, spherical, and adorned with two circular signa composed of small sclerotized dots.

Of special interest in the males are the coremata (Figs. 10-12). These brushes of hairs, white in dissected dried specimens, are located in an invaginated pocket between the seventh and eighth abdominal sterna and are presumably scent-distributing organs, for a liquid is associated with the coremata in live specimens (Lane, 1957). MacNeill (1962) has reported a voluntary display of coremata in *Estigmene acrea* (Drury), but we are unaware of any published accounts of their function. Lane (1957) has reported on the coremata in a live European *P. fuliginosa*, while numerous other observations have been made on other arctiids (see MacNeill, 1962, for a good bibliography, to which may be added Siewers, 1879, and Kershaw, 1953).

All three North American species have a "striated band," a term used by Forbes and Franclémont (1958) and Forbes (1960) for a series of curved, parallel grooves on the anterior edge of each metepisternum, in males and females alike (Figs. 16-18). Blest et al. (1963) referred to the swollen, grooved metepisternal sclerite of the arctiid *Melese laodamia* Druce as a "tymbal organ." They found that vertical compression and relaxation of the sclerite caused it to buckle inward and outward, respectively, producing a series of clicks as the 15-20 grooves rubbed against one another. While not precluding the possibil-
ity of a variety of functions in different species, they admitted the pos-
sibility that the clicking sounds produced by the tymbal organ could
confuse the echolocation system of bats, for the sounds of bats and
this moth were similar. Later, Dunning and Roeder (1965), working
with the arctids *Hyalisidota tessellaris* (Smith & Abbott) and *Pyrrharch-
tia isabella* (S. & A.), found that ultrasonic sounds emitted by the "mi-
crotymbal organs" on the metepisterna could provide the moths with
some degree of protection from bats.

The venation (Fig. 19) of all three species is subject to much
variation, although the general plan follows that of *Spilosoma* or *Estig-
mene*. Dyar (1891) was correct in pointing out that the origin of R₅ is
before that of R₄ in the forewing, and not the converse as figured by
Smith (1890: 235, fig. 8)—although Smith may have examined an aber-
rant specimen. The major sources of variation are the points at which
R₁ and R₅ arise (sometimes very close to the apex of the cell, other
times not); the point of origin of R₆ when compared to the origins of R₅
and R₆; the distances between the points of origin of M₁, M₂, and
Cu₁ in both wings; and whether R₅ and M₃ of the hindwing both arise
from the apex of the cell, or whether they have a common stalk, which
may be either short or long. Several interesting venational extremes
were noted: R₅ and R₆ of the forewing diverging from the same point
(two specimens of *P. juliginosa*); a forked R₅ vein, resulting in a six-
branched radius (one *P. assimilans*); and R₅ arising before the apex of
the cell (*i.e.*, before the origins of R₃₋₅; seen in one specimen of *P.
assimilans*).

Because of the sexually dimorphic shape of the forewing, R₅ in
females extends slightly beyond the end of M₁ (with the inner margin
horizontal), while in males R₅, M₁, M₂, and M₃ terminate about
equidistant from the base of the wing. "In other's" words, the forewing
of females is produced at the apex, while the outer margin of the male
forewing is nearly perpendicular to the inner margin.

The drawings in this paper were made on No. 2 Rossboard with
the aid of a grid eyepiece and graph paper. The scale of the drawings
varies, but in all cases similar structures (*e.g.*, valvae) are drawn
to the same scale, thus permitting comparisons of size.

The solid black circles on the distribution maps represent the
approximate collection sites of specimens examined. Triangles repres-
ent either published records we believe to be reliable (Brodie, 1929;
Ferguson, 1953; Jones, 1951; Krogerus, 1954) or records obtained
from individuals (Richard W. Holzman, Michigan; J. C. E. Riotte, On-
tario). The small open circles represent major cities. The distribu-
tion maps were prepared on map number 302, North America, pub-
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The capitalized color names in the descriptions refer to the
standardized color names in Ridgway (1912).

In the list of specimens examined following the discussion of
each species, states, counties, and cities are arranged alphabetically.
Following each citation is the place of deposition, according to the
abbreviations below.
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AMNH American Museum of Natural History, New York (Frederick H. Rindge).
ANS Academy of Natural Sciences, Philadelphia (the late Harold J. Grant, Jr.).
BM British Museum (Natural History), London (Allan Watson).
CAS California Academy of Sciences, San Francisco (C. Don MacNeill).
CNC Canadian National Collection, Ottawa (D. F. Hardwick).
CNHM Chicago Natural History Museum (Alex K. Wyatt).
CU Cornell University, Ithaca, New York (L. L. Pechuman).
FRLA Forest Research Laboratory, Calgary, Alberta (H. A. Tripp).
FRLM Forest Research Laboratory, Winnipeg, Manitoba (K. R. Elliot).
FRLNB Forest Research Laboratory, Fredericton, New Brunswick (R. S. Forbes).
FRLQ Forest Research Laboratory, Sillery, Quebec (R. Martineau).
INHS Illinois Natural History Survey, Urbana (Wallace E. LaBerge).
KU University of Kansas, Lawrence (George W. Byers).
MASS University of Massachusetts, Amherst (Marion E. Smith).
MCZ Museum of Comparative Zoology, Harvard University, Cambridge, Massachusetts (Howard E. Evans).
MINN University of Minnesota, St. Paul (Frederick W. Stehr).
MSU Michigan State University, East Lansing (Roland L. Fischer).
NDSU North Dakota State University, Fargo (R. L. Post).
ODA Oregon Department of Agriculture, Salem (Kenneth Goeden).
OSM Ohio State Museum, Columbus (Edward S. Thomas).
OSU Oregon State University, Corvallis (Jack Lattin).
PM Peabody Museum of Natural History, Yale University, New Haven, Connecticut (Douglas C. Ferguson).
UMMZ University of Michigan Museum of Zoology, Ann Arbor (Thomas E. Moore).
UNH University of New Hampshire, Durham (J. G. Conklin, Wallace J. Morse).

The following persons kindly allowed us to examine specimens in their personal collections. Their cooperation in this project is greatly appreciated.

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KEY TO MALE NORTH AMERICAN PHRAGMATOBIA

1. Valva with two mesad-directed projections about midway from base to apex, one arising from dorsal margin, the other from the ventral margin (Fig. 15); striated band on metepisternum well-developed (Fig. 18); forewing with no indication of dark antemedial or postmedial lines (Figs. 30-33); coremata well-developed; forewing 14-15mm from base to apex . . . . . . . .

Mesal face of valva with a single, central, spade-like projection (Figs. 13, 14); striated band variable; forewing usually with at least traces of antemedial and/or postmedial lines (Figs. 26-29, 34)–if no lines on forewing, then forewing over 17mm and costal margin of hindwing not blackened; coremata variable . . 2
2. Distal end of aedeagus with a sclerotized dorsal "comb" (Fig. 7); striated band on metepisternum well-developed (Fig. 16); hindwing costal margin blackened; wings translucent; coremata well-developed; forewing 15-16mm ............... P. lineata, NEW SPECIES (p. 40)

Distal end of aedeagus with at most a few lateral teeth (Fig. 8); striated band poorly-developed, may appear absent (Fig. 17); hindwing costal margin not blackened, concolorous with discal area; wings opaque; coremata small and poorly-developed; forewing usually over 17mm ............... P. assimilans (p. 57)

KEY TO FEMALE NORTH AMERICAN PHRAGMATOBIA

1. Striated band on metepisternum poorly-developed, may appear absent (Fig. 17); costal margin of hindwing not suffused with black, concolorous with pink discal area; wings opaque; forewing usually with at least traces of antemedial and/or postmedial lines (Figs. 27, 29); mostly boreal in distribution ................. P. assimilans (p. 57)

Striated band well-developed (Figs. 16, 18); costal margin of hindwing suffused with distinct but varying degrees of black, usually invading at least part of the discal cell; wings, especially hindwing, translucent; forewing lines present or absent .......... 2

2. Always a trace of at least one of the forewing lines (antemedial, postmedial, subterminal, adterminal—Fig. 35); striated band usually with more than 9 ridges (Fig. 16); known from southern Alberta and Manitoba, and south of a line drawn from central Maine to northern N.Y., extreme southern Ontario, southern Mich., west to eastern Nebraska (Fig. 38) ................. P. lineata, NEW SPECIES (p. 40)

Never a trace of dark lines on the forewing, although an obscure fragmentary subterminal smudge may be visible near the apex of the forewing (Figs. 31, 33); striated band usually with 7 to 9 ridges (Fig. 18); range includes that of former, plus Colo., Cal., Pacific Northwest and throughout Canada and Alaska north to the tundra (Fig. 36) ............... P. fuliginosa rubricosa (p. 49)

PHRAGMATOBIA LINEATA Newman and Donahue, NEW SPECIES

(Figs. 1, 4, 7, 10, 13, 16, 20, 23, 34, 35)

DIAGNOSIS: In some cases it is necessary to examine the male genitalia for an infallible identification, but in all except the most badly rubbed specimens the examination of two pattern characters will be adequate for identification: the presence of any lines on the forewing, and the continuance of the marginal (or submarginal) black band on the hindwing upperside along the costal margin. The first character separates P. lineata from P. fuliginosa, the second character separates P. lineata from P. assimilans.
This species has long been confused with both *P. fuliginosa* and *P. assimilans*, because it has some characters of each. Wherever *P. lineata* occurs, *P. fuliginosa* also occurs, the two being active at the same time. This sympatry and synchrony, and the similarity of size and wing translucency, has led previous workers to assume that *P. lineata* was a "form" of *P. fuliginosa*. On the other hand, *P. lineata* resembles *P. assimilans* in that both have lines on the forewing (absent from *P. fuliginosa*), and both have a single projection on the mesal face of the valva (two projections in *P. fuliginosa*).

Females are more melanic than males, but always have traces of lines on the forewing (separating them from *P. fuliginosa* females). They can be distinguished from similar females of *P. assimilans* by the shiny, translucent hindwing and the dark costal margin on the hindwing upperside. Differences in the female genitalia are slight. Further differences among the three species are emphasized in the key.

**HOLOTYPE MALE (Fig. 34)**

**HE A D**: small, antennae ventrally ciliate, dorsally with appressed white scales. Vertex, frons, and palpi with spreading Mahogany Red hairs (appearing Brazil Red under strong light and magnification). Eyes small, hairless.

**THORAX**: vestiture dense, dorsally Mahogany Red, ventrally Chestnut (English Red under light and magnification). Femora with dorsally appressed Rufous hairs and ventrally spreading Mahogany Red hairs, except for a dorso-distal blackish patch ("knee"). Tibiae with appressed Rufous and Mahogany Red hairs and scales. Tarsi almost entirely Slate. Mid-tibiae with one pair of whitish apical spurs, hind-tibiae with two pairs of whitish spurs. Metepisternum as in Figure 16.

Forewing Upperside: scales sparse and semi-erect which, coupled with transparent membrane, makes wings translucent. Because of this translucency the general color is hard to define, but it is near Kaiser Brown, becoming slightly darker at the base. Blackish markings as follows: an antemedial smudge immediately beneath discal cell, just proximad of origin of vein Cu₂; a spot at upper end of discal cell, and a larger one at lower end of discal cell; an irregular postmedial line, about 1mm wide, arising on inner margin 3/4 of distance between base of wing and distal end of vein 2A, narrowing to 0.5mm and more or less directed towards apex as far as vein M₃, then curving proximad and continuing faintly and perpendicularly to a point on the radial sector about one-third the distance between the upper cell spot and the apex; a small, subapical mark between veins R₄ and R₅, a straight subapical bar from the apex to vein M₂, directed to a point about 1mm distad of the origin of the postmedial line; an antermal line parallel to the outer margin, extending from apex to tornus; and a narrow terminal line (the dark bases of the fringe scales). Fring Coral Pink, extending from apex to tornus, becoming short dark hairs on inner margin, increasing in length to the base of the wing, where there is a tuft of Coral Pink hairs with Mahogany Red tips. Veins concolorous with scales.
Forewing Underside: shiny Coral Pink; the two cell spots repeated, other dark dorsal markings showing through from upperside; costal margin appearing darker because of denser arrangement of scales; frenulum simple, stout; retinaculum Slate colored; very thinly scaled except for a tuft of appressed Coral Pink hairs arising at base of cell Cu₂ and directed anteriorly to retinaculum; a small ovoid patch devoid of scales at base of anal cell.

Hindwing Upperside: Coral Pink throughout, except: a blackish terminal band beginning 0.7mm posterior of distal end of vein 2A, where it is 1.2mm wide, becoming wider towards the apex, where it is 2.6mm wide, then continuing along costal margin to a point about 1/3 the distance between base and apex; a blackish patch on the M₁-M₂ crossvein at end of discal cell, conjoined to the blackish costal margin; and a small blackish spot at lower end of discal cell. Very thinly scaled and shiny in the discal area, with Coral Pink hairs becoming progressively longer and more dense from the posterior side of the discal cell to the inner margin; a dense patch of Coral Pink scales in basal third of costal margin, coinciding with the bare patch on inner margin of forewing underside; fringe Coral Pink.

Hindwing Underside: very thinly scaled and shiny Coral Pink, except for the two spots at the end of the cell, as on upperside, with the marginal blackish line showing through from upperside. Costal margin more densely scaled.

A B D O M E N: dorsum Coral Red, with a median row of 6 black spots, and a lateral row of 6 black spots on each side; venter, below the lateral rows of spots, Chestnut.

Coremata (Fig. 10), described from a topotypic paratype, 17 Aug. 1964: four well-developed brushes of soft white (in dissected dried specimens) hairs in two contiguous pouches invaginated between the seventh and eighth sternum.

Genitalia (Figs. 1, 4, 7, 13), described from a topotypic paratype, 17 Aug. 1964: Uncus (Fig. 1) long and narrow; valvae (Fig. 13) long, simple, with a single spade-like projection on the mesal face, about midway between base and apex; aedeagus (Fig. 7) stout, with a broad sclerotized dorsal "comb" arising from the right side on the distal end, and a long, toothed, strap-like sclerotized area arising from the left side, the latter extended posteriorly and attached to the vesica; vesica adorned with sclerotized thorn-like cornuti of varying size.

SIZE: forewing 15mm from base to apex (end of vein R₄); 12mm from base to tornus (end of vein 2A); 8mm from apex to tornus (straight line). Hindwing 12mm from base to apex (end of vein R₄); 6mm from base to anal angle (end of vein 3A); 10mm from apex to anal angle (straight line).

ALLOTYPE FEMALE (Fig. 35)

Darker red and more melanic than male.
HEAD: as in holotype.

THORAX: vestiture as in male, except that ventral hairs are darker (Brazil Red), and dorsal appressed hairs on femur are darker (Flame Scarlet). Legs otherwise as in holotype male.

Forewing Upperside: darker and not so translucent as in holotype male, but scales still semi-erect; ground color appears near Carob Brown. The blackish markings, slightly different from those of the holotype male, are as follows: a faint antemedial line, beginning at midpoint of posterior edge of discal cell, continuing obliquely proximad to inner margin; one spot each at upper and lower ends of apex of discal cell, more or less confluent and outlining end of the cell; a postmedial line as in male, but so faint it is represented by slightly darker scales visible only under magnification; a Y-shaped subterminal line, arising from the inner margin near end of vein 2A and proceeding, more or less parallel to the outer margin, to cell M₂, where it branches, the distal branch extending straight to apex, becoming wider from vein R₅ to apex, the proximal branch extending straight to a point on costa about midway between end of cell and apex of wing, widening from cell R₅ to costa; and a microscopic terminal line, formed by the dark bases of the fringe scales. Fringe Begonia Rose from apex to tornus, then continuing as small hairs, concolorous with ground color, along inner margin to base, where the hairs become longer, more dense, and Rufous.

Forewing Underside: as in holotype, except darker (Begonia Rose), blackish dorsal markings repeated by slightly darker scales; frenulum compound, subcostal retinaculum absent.

Hindwing Upperside: blackish, except for Begonia Rose fringe, basal third of costal margin, and anal area. Marginal black band wide, diffuse, becoming paler and suffusing proximad to cover the entire wing except: the patch of Begonia Rose scales in the basal third of the costal margin (as in male); the Begonia Rose fringe from apex to inner margin; and the Begonia Rose hairs from the basal two-thirds of midcell Cu₂ to inner margin, which become progressively longer towards inner margin. Scattered pink scales in the discal area make that area more diffusely black than the marginal area. Distal end of discal cell outlined in black, slightly darker than ground color, but not present as distinct spots as in holotype.

Hindwing Underside: as in holotype male, but darker (Begonia Rose), except for a 2mm-wide submarginal band that repeats the dorsal marginal band. Discal and marginal areas with scattered Begonia Rose scales, becoming more dense at base of costal margin and in space between discal cell and inner margin. Subcostal vein with a row of blackish scales.

ABDOMEN: dorsum spotted as in male, but ground color darker (Rose Red); venter Brazil Red.

Genitalia (Figs. 20, 23), described from a topotypic paratype, 1 July 1965: ductus bursae very wide, inflated, heavily sclerotized;
cervix bursae large, rubbery, somewhat convoluted; corpus bursae large, spherical, membranous, with two round signa of sclerotized dots, one signum on each side. Lamella antevaginalis well-sclerotized, expanded laterally; lamella postvaginalis irregular in shape, with a dense patch of fine hair on the dorsal side. The sclerotized plates to which the apophyses anteriores are attached are broad and rounded. Other structures as illustrated.

**SIZE:** forewing 15mm from base to apex (end of vein R₁); 10mm from base to tornus (end of vein 2A); 8mm from apex to tornus (straight line). Hindwing 10.5mm from base to apex (end of vein R₁); 6mm from base to anal angle (end of vein 3A); 9.5mm from apex to anal angle (straight line).

**VARIATION**

As with many species of arctiids, the variation in this species is considerable. In the long series of paratypes the following individual variation has been observed (the individual variation is so great that geographical variation, if any, would be concealed to all but detailed statistical analyses):

**MALES:** general coloration is fairly constant, although the wings of some specimens may be less densely scaled than others, so that light reflecting from the membrane gives the hindwings a more shiny, rosy appearance. Grease may cause the thorax and abdomen to darken and lose their color.

Forewing Pattern (upperside): at least one line is always distinct (usually the postmedial). Many specimens have more prominent lines than those described in the holotype. The smudge beneath the cell in the holotype is developed in other specimens to a conspicuous, oblique antemedial line, extending from the posterior side of the cell to the inner margin (as described in the allotype). The postmedial line is constant in configuration but variable in intensity. The subterminal line, when well-developed, is Y-shaped, as described in the allotype. There is often an additional, adterminal, line between veins M₁ and Cu₂. The basic pattern elements, then, are: an oblique antemedial line from the posterior side of the discal cell to the inner margin; a curved postmedial line; a Y-shaped subterminal line; and an adterminal line. Very rarely the antemedial line is bent at the posterior margin of the discal cell, and continues proximad across the cell, as in well-marked specimens of *P. assimilans*. The result is an excurved antemedial line. The variability of forewing markings in this species is a result of the permutations, combinations, and degree of development of these four pattern elements and their parts.

Hindwing Pattern (upperside): the blackish band on the hindwing may be marginal, as in the holotype, or it may be submarginal, with the pink ground color occurring between the band and the fringe. When the band is submarginal it usually touches the apex and anal angle, diverging most from the outer margin between veins M₁ and Cu₂. This
band is almost always continuous and fairly uniform, but in some specimens it becomes constricted to form a series of confluent maculae, occasionally leaving an isolated spot or two near the anal angle.

**Females:** the general coloration is fairly constant, although the shade of pink on the hindwing is subject to some variation, perhaps partly due to age of the specimen.

Forewing Pattern (upperside): because the ground color is darker than in males, it is more difficult to distinguish the pattern. But all four pattern elements are present in well-marked specimens, and these elements undergo variation as described for males.

Hindwing Pattern (upperside): the extent of black suffusion varies, but it always extends to the outer margin (not becoming macular or submarginal as in males), and nearly always enters part of the discal cell. Consequently, the extent of pink also varies. The anal area is always conspicuously pink or red.

**LIFE HISTORY**

*Phragmatobia lineata* is apparently at least double brooded, and may be triple brooded. Light trap captures at the type locality in 1965 showed three definite flight periods: early May (two males); late June to late July (50 males, five females); and mid-August to mid-September (32 males, one female). Small numbers of specimens have been recorded from other localities from early April to early October.

Although we have not examined any larvae of this species, we do have a good idea of its general appearance. Goodhue (1902) reared this species in Webster, New Hampshire, but thought he was dealing with either *P. assimilans* var. "franconia" or an undescribed species. He described the larva as "... pale yellow, thickly covered with rather long soft light yellow hair" (p. 286), adding that it looked exactly like the larva of *Euchaetias oregonensis*. We have examined two male *P. lineata* from Webster (in coll. Brower and UNH) with pin labels that verify the above description. Although the larval descriptions for *P. fuliginosa* and *P. assimilans* are conflicting, none of them resembles this description of the *P. lineata* larva. Overwintering is in the larval or pupal stages, or perhaps both.

**FOODPLANTS:** *Eupatorium* (thoroughwort, Joe-Pye weed, etc.) (Goodhue, 1902), and probably on many other low herbs, such as *Solidago* (goldenrod), *Taraxacum* (dandelion), and *Rumex* (dock).

**FIGURES:** a very good illustration of a *P. lineata* female appears in Beutenmüller (1898: pl. 16, fig. 6, as *P. fuliginosa*), while Hampson (1901: fig. 135, p. 244) depicts a *P. lineata*, probably a female, as *P. fuliginosa c*. Although Hampson's figure clearly shows the postmedial line and part of the subterminal line, his description omits mention of them.
DISTRIBUTION
(Fig. 38)

Calgary, Alberta ( ) and southern Manitoba; and in the Transition and Upper Austral Zones, usually in deciduous forest areas, from eastern Nebraska, Iowa, and northeastern Missouri east through northern Illinois, southern Michigan, extreme southwestern Ontario, northern New York and central Maine, south to northern Indiana, northern Ohio, Pennsylvania, and central New Jersey. The northern limit of distribution is hard to define, but the known southern limit is near 40° north latitude. The Alberta and Manitoba records are exceptional, in that they are widely disjunct from other populations. Further collecting may prove these populations to be continuous with those in the United States.

TYPES

The holotype male was collected by black light trap in an area surrounded by uncultivated fields one mile east of Galien, Berrien County, Michigan (Township 8 South, Range 19 West, Section 1) on 2 July 1965 by R.E. Thompson. The allotype female was collected by R.E. Thompson at the same locality on 23 June 1965. Both the holotype and allotype are deposited in the Entomology Museum at Michigan State University. Paratypes are as follows (all specimens, except a disintegrated male from Van Buren Co., Mich., are designated as paratypes):

SPECMENS EXAMINED (526)


MISSOURI. PIKE: Louisiana, 23 Sept. 1905, 1♂ (CAS).


NEW HAMPSHIRE. no data, "Bred. Larva dark yellow, thickly covered with short stiff bristles, after the manner of A. isabella," 1♀ (UNH); "E. Light," 2♂ (UNH); White Mountains, 2♂ (MSU). MERRIMACK: Webster, W. F. Fiske, 1♀ (UNH);


OHIO. PAULDING: Sec. 15, Benton Township, 8 Sept. 1963, Homer Price, 1♂ (Price).


CANADA

ALBERTA: Calgary, Head of Pine Creek, 1 & 7 May 1914, 27 May 1894, F. H. Wolley Dod, 2♂, 1♀ (CNC); Calgary, "at rest in town," 30 May 1907, F. H. Wolley Dod, 1♀ (CNC).


PHRAGMATOBIA FULIGINOSA RUBRICOSA (Harris)

(Figs. 3, 6, 9, 12, 15, 18, 22, 25, 30-33)

Arctia fuliginosa Linnaeus, 1758: 509.

Phragmatobia fuliginosa, Stephens, 1829: 74.

Arctia rubricosa Harris, 1841: 253 (nec. P. rubricosta Dognin, 1889: 193, type locality Loja, Ecuador).

Phragmatobia fuliginosa rubricosa, Rothschild, 1910: 116;

McDunnough, 1938: 49.

The infraspecific names proposed for this species in Europe, several of which are listed in the introduction, are not considered by us to be applicable to this subspecies.

Phragmatobia rubricosa was generally accepted as a valid species distinct from P. fuliginosa until Dyar (1891) synonymized the two. Allan Watson of the British Museum (Natural History) concurs with us (pers. comm.) in believing that the North American population is at least subspecifically distinct from the named European populations.

This judgment is based primarily on color and pattern differences: most European specimens we have seen are much lighter in color, and in most the hindwing band is more or less macular, especially towards the anal angle—a character seldom seen in North American specimens. There are also minor differences in the male genitalia, especially in the vesica, that may prove to be valid criteria for the specific separation of the two populations, but this decision must await further work on the Eurasian species.

Phragmatobia fuliginosa rubricosa is closely allied to P. lineata, and has been confused with it for years. Any published references to a "form" with lines on the forewing (as in Forbes, 1960, and Beutenmüller, 1898) refer to P. lineata only, for P. fuliginosa never has lines on the forewing (with two notable exceptions: a male in the A.E. Brower collection from Dennistown, Somerset Co., Maine, 21 July, and a male in the Hessel collection from Washington, Litchfield Co., Conn., 22 Sept. 1961, but the genitalia proved them to be P. fuliginosa—the forewings are marked as in P. lineata, the hindwings are dark as in P. fuliginosa).

While the absence of forewing lines separates this species from P. lineata, the shiny, translucent wings, the smaller size, the dark costal margin of the hindwing upperside, and the flying time (usually later in the season) serve to separate this species from P. assimilans. A few unusually small specimens of P. fuliginosa rubricosa occur in May and early June (the flying time of P. assimilans), but the great majority is collected from late June through September.

Whenever the identity of a specimen is uncertain, an examination of the male genitalia quickly dispels any doubt. A pair of jeweler's forceps, a fine brush, and a dissecting microscope are the only tools needed to remove the anal hairs and examine the exposed valvae. This is the only North American species with two projections on the valva (Fig. 15).
MALES tend to be slightly smaller than males of *P. lineata*. The general appearance is much the same, except that lines on the forewing are absent, and the hindwing is usually more suffused with black. The distal edge of the black band on the hindwing upperside usually extends to the outer margin, while the proximal edge usually suffused inward to the discal area, sometimes (Fig. 32) leaving only the costal patch, fringe, and anal area pink or red, as in females. In those few specimens in which the black band is submarginal (Fig. 30), the band is usually more sharply defined, distally dentate, and tends to be broken up into spots, as in most European specimens. This variation, occurring throughout the North American range of this species, appears to be individual rather than geographic. The coremata (Fig. 12) are well-developed, as in *P. lineata*.

FEMALES are about the same size as *P. fuliginosa* males, but are generally smaller than *P. lineata* females. In general appearance they are much darker than males, with the black on the hindwing always extending to the outer margin (Figs. 31, 33).

GENITALIA: The male genitalia (Figs. 3, 6, 9, 15) are distinctive. In the absence of adequate series of specimens of known parentage, no constant, outstanding differences in female genitalia have been noted at this time. Figures 22 and 25 will, however, give an idea of some potential specific differences.

LIFE HISTORY: The larvae are apparently quite variable in color, judging from published descriptions (Beutemüller, 1898; Edwards, 1887; Edwards & Elliot, 1883; Goodhue, 1902). The descriptions agree in that the skin of the larva is black or slate, but the color of the setae apparently varies from chestnut to a foxy yellow. Some authors say the larvae have pale yellow dorsal and lateral lines, but we have not seen specimens marked in this manner. According to Goodhue (1902), the thoracic setae are usually darker.

There are probably at least two broods a year. We have seen larvae crawling about in the autumn looking for places to overwinter, in a manner similar to that of the common banded woolly bear (*Pyrarctia isabella*). On 27 February 1966 Lois Donahue found such an overwintering larva under a log beside an old field in Livingston Co., Michigan. When brought indoors it refused to eat dandelion and pupated in a flimsy cocoon on 2 March 1966, emerging 10 days later.

Specimen labels, and our own observations, show that some larvae pupate in the fall, and overwinter as pupae. The few early spring specimens that have been collected, especially in boreal localities, probably represent pupae or ultimate-instar larvae that overwintered, while most specimens flying later in the spring and in early summer probably are the offspring of the spring brood. Larvae which overwintered in earlier instars may produce moths that fly later, intermingling with the early spring brood. These conjectural statements, based on meager observations and a few notes on specimen labels, only hint at what is actually happening. The dynamics of the various broods can only be determined after further study.
PARASITES: *Zenillia affinis* (Fallén) (Diptera: Tachinidae) has been reared from the larvae of what was identified as *Phragmatobia fuliginosa rubricosa* (Schaffner & Griswold, 1934). The fly referred to is possibly *Hubneria estigmenens* (Sellers)—a species with which Fallén's species has been confused.

FOODPLANTS: The larvae are general feeders on low herbaceous vegetation. Specific records include *Solidago* (goldenrod), *Eupatorium* (thoroughwort, Joe-Pye weed, etc.), *Symlocarpus* (skunk cabbage), *Vernonia noveboracensis* (ironweed), *Rumex* (dock), *Plantago* (plantain), *Myrica gale* (sweet gale), and *Helianthus* (sunflower) (Beutenmüller, 1898; Dirks, 1937; Edwards & Elliot, 1883; Edwards, 1889; Jones, 1951; and specimen labels).

FIGURES: This species has been correctly figured in Holland (1903: fig. 70; pl. 14, no. 31) and Seitz (1919: pl. 38i, as *rubricosa*). For figures erroneously referring to this species, see the discussion under *P. lineata*.

TYPE LOCALITY: Unknown, but presumed to be somewhere in Massachusetts. The original description does not designate a type locality, but Harris (1841), writing about Massachusetts insects, says "This moth is rare; and it appears here in July and August" (p. 253), the "here" assumed to mean Massachusetts.

TYPE SPECIMEN: Male (genitalia examined), Type No. 26372, in the Harris Collection at the Museum of Comparative Zoology, Harvard University, Cambridge, Massachusetts.

DISTRIBUTION (Fig. 36): South of the tundra from central Alaska to Newfoundland, south to California (Boisduval, 1852, in addition to the two specimens cited here), Montana, and Colorado, and south to North Dakota, Minnesota, northern Illinois, northern Indiana, southern Michigan, Pennsylvania, and New Jersey. Probably also occurs in central British Columbia, Alberta, at high elevations all along the coastal ranges and Rockies, and in Ohio. The few specimens that have been collected in montane or far-northern areas were caught during the daytime, when the moths were active. Holland (1903) states that this species occurs south along the Appalachians to the Carolinas, and Seitz (1919) says it occurs in "Carolina" (possibly repeating Holland), but we have neither seen nor heard of any specimens from south of Pennsylvania. Rothschild's (1910) record of Florida specimens must be viewed with extreme doubt.

SPECIMENS EXAMINED (991)


COLORADO. no data: Smith, 2♂ (BM); strecker coll., 1♂ (CNHM). DENVER: Denver, Engel coll., 2♂ (CM); 8-19 Aug., Barnes coll., 3♂, 1♀ (no date) (USNM). GARFIELD: Glenwood Springs, Barnes coll., 1♂ (USNM). UNKNOWN LOCALITIES: "Colorado, Bruce," 2♂, 1♀ (no date), Edward T. Owen (USNM); Hayden Mts., Oslar, Aug., Barnes coll., 1♂ (USNM) [There is a Hayden Peak somewhere in NW Colorado, while there is an Oslar in Ouray Co.; J. Oslar, however, may have been the collector.]


MASSACHUSETTS. no data: 3♂ (INHS); 17 July 1901, J. G. Grundell, 1♂ (CAS); also two males in the Harris collection, MCZ, one of which is type of Arctia rubricorsa, both presumably from Mass. BARNSTABLE: Barnstable, 20 July 1952, C. P. Kimball, 1♂ (AMNH). BERKSHIRE: Tyringham, 16-27 July 1953, A. E. Treat, 6♂ (AMNH). DUKES: Marthas Vineyard Is., 29 July & 8 Aug., Frank M. Jones, 1♂, 1♀ (PM). HAMPSHIRE: Amherst, 4 & 31 July 1964, R. W. Koss, 1♂, 1♀ (MSU); 27 April 1938, Savage, 1♂ (MASS); 22 May 1948, J. O'Neil, 1♀ (MASS); 27 July 1942, S. K. & A. L., 1♂ (MASS); 23 Aug. 1938, W. Kulash, 1♂ (MASS); 20-31 July 1935, 1944, Marion E. Smith, 4♂ (MCZ). MIDDLESEX: Natiek, 1♂ (MCZ). Sherborn, 22 July 1900,
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CANADA


NEWFOUNDLAND. Harmon Field (air base, at Stephenville), 16 June 1949, F.G. DiLabio, 1♂ (CNC).


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PHRAGMATOBIA ASSIMILANS Walker

(Figs. 2, 5, 8, 11, 14, 17, 21, 24, 26-29)

Phragmatobia assimilans Walker, 1855: 630.
Phragmatobia var. franconia Slosson, 1891a: 3.
Phragmatobia assimilans var. franconia Slosson, 1891b: 41, pl. 3, fig. 2 (nec. franconica, franconiae, auctores, in error).

This variable species, our largest Phragmatobia, was unknown in United States collections until Annie Slosson (1889, 1890) collected it in northern New Hampshire. Because the taxonomists of the latter half of the nineteenth century had no comparative material of this rarely-collected species, they assumed it to be a synonym of P. fuliginosa rubricosa. Even after its rediscovery, Kirby (1892: 245, 911) persisted in retaining P. assimilans as a synonym of P. rubricosa.

The similarity of this species to P. lineata has caused the two to be confused in many collections. Indeed, P. assimilans appears to be more closely related to P. lineata than to P. fuliginosa (based on similarity of forewing pattern and valvae), but P. assimilans can be easily separated from P. lineata by the following characters: opaque wings, larger size, more boreal distribution (the two being sympatric, so far as is known, only in Calgary, Alberta, southern Manitoba, eastern New York, and New England), absence of a blackened costal margin on the hindwing upperside, the early flight period (May and June), and the absence of the sclerotized "comb" on the distal end of the aedeagus.

P. assimilans can similarly be distinguished from P. fuliginosa rubricosa by most of the above characters (opaque wings, larger size, absence of blackened costal margin on the hindwing upperside), and also by the distinctive genitalia.

MALES are robust, "hairy" moths subject to a great deal of variation. Most specimens are strongly marked and have black lines on the forewing (Fig. 28), as in P. lineata: an excurred antemedial line directed obliquely distad from the inner margin to the posterior side of the discal cell, then proceeding obliquely proximad to the anterior side of the discal cell (in P. lineata this line only rarely crosses the cell); an excurred postmedial line; a curved subterminal line, occasionally Y-shaped, with the distal arm of the Y going to the apex; and an adterm-
inal line. Some (or, rarely, all) of these pattern elements may be ab-
sent (Fig. 26), while the intensity of various parts of the elements is
extremely variable. Usually, however, the antemedial and postmedial
lines are well-developed and conspicuous.

The ground color of the forewing varies from light to dark
brown, with variable amounts of black "pepperin." The fringe is pink.
The black band on the hindwing upperside is also subject to
great variation. In those specimens with a dark forewing the band on
the hind wing is broad, uniform, and marginal (Fig. 28). In lighter
specimens the band becomes submarginal (Fig. 26), and tends to be
macular and broken up into an anterior segment and a posterior seg-
ment, or is reduced to one or two irregular black spots near the anal
angle, or, very rarely, the hindwing band is completely absent (only
one such specimen seen—one of Annie Slosson's from Franconia, N.H.,
in the American Museum of Natural History, perhaps the one she fig-
ured, 1891b, pl. 3, fig. 1).

The hindwing fringe, costa, and discal area are varying shades
of bright rosy pink.
The coremata (Fig. 11) are poorly developed.

FEMALES (Figs. 27, 29) are relatively rare in collections—
most specimens appear to have been reared. They come to lights
only rarely, if at all—a large number of males have been collected in
Michigan and Nova Scotia, for example, but no females have ever been
collected from these areas. Females tend to be darker than males,
and almost always have some part of the forewing pattern well de-
veloped. The black band on the hindwing is broad, and extends to the out-
er margin in all specimens we have examined.

SYNONYMY: The variety "franconia" described by Slosson
(1891a; 1891b, pl. 3, fig. 2) is merely a well-marked specimen near
one end of a broad range of individual variation, although it is marked-
ly different from the other, immaculate specimen she had before her.
Obviously, "franconia" is not a variety at all, and is merely a synonym
of P. assimilans. The type of "franconia"—a male, not a female as
stated in the original description—is deposited in the American Muse-
um of Natural History. The genitalia were examined to eliminate any
doubt concerning the identity of the specimen.

GENITALIA: Surprisingly, the male genitalia of this large spe-
cies are smaller than the genitalia of the other two species. The sin-
gle projection on the mesal face of the valva (Fig. 14) allies this spe-
cies to P. lineata, but the absence of the "comb" on the distal end of
the aedeagus (Fig. 8) identifies P. assimilans. The female genitalia
(Figs. 21, 24) are patterned after the basic plan of the other species,
but are smaller. The lamella antevaginalis is poorly developed, and
the sclerotized plate to which the apophysis anteriore is attached is
quadrate. As with the other species of Phragmatobia, the taxonomic
value of the female genitalic structures can only be determined after
further work.
LIFE HISTORY: *Phragmatobia assimilans* appears to have only one generation per year. The adults are on the wing from mid-April to mid-June, depending on the locality. The only exceptions to this early flight period are represented by a series of 18 specimens from Constance Bay, Carleton Co., Ontario, collected from 19 July to 2 August 1935 by G.S. Walley; and two females from Edmonton, Alberta, collected on 5 & 6 August 1917. These specimens may represent a second brood, or reared specimens whose normal developmental period was affected, or they may have been mis-labeled.

The larvae overwinter in the last instar, and the following spring they crawl about looking for a place to pupate. The hardiness of these larvae is remarkable—they may be seen crawling about when the ambient temperature is near freezing, and have even been seen crawling on snow (Gibson, 1911). Data on specimen labels indicate that pupation takes place in a flimsy cocoon constructed off the ground in trees or brush.

The larval stages have been described by Gibson (1911), so we will only point out that the larvae would appear similar to larvae of *P. fuliginosa*, but would be darker (mouse gray), without any trace of dorsal or lateral markings. Note that the larval description by Goodhue (1902) applies to *P. lineata*, not *P. assimilans*.

PARASITES: *Exorista cheloniae* (Rondani) (Gibson, 1911)—probably a species of *Carcelia*, for this species is not Nearctic (Diptera: Tachinidae).

FOODPLANTS: *Mertensia*? (bluebells), reared on *Taraxacum* (dandelion) and *Plantago* (plantain) by Gibson (1911); *Betula papyrifera* (white birch) (McGegan, 1958); *Rubus* (raspberry, blackberry), specimen label, Forest Research Laboratory, Manitoba; *Populus balsamifera* (balsam poplar), specimen label, Forest Research Laboratory, Alberta.

FIGURES: *Phragmatobia assimilans* has been properly figured by Ferguson (1953: pl. 3, fig. 8); Hampson (1901: pl. 43, fig. 22); Seitz (1919: pl. 38, row i, as *assimilans* and *franconia*); and Slosson (1891b: pl. 3, fig. 1, as *assimilans*, and fig. 2, as var. *franconia*).

TYPE LOCALITY: The original description (Walker, 1855: 630) says only that the specimens were from the "United States," and were "Presented by E. Doubleday, Esq.," while the undated labels on the two syntypes say "U.S. America" (Allan Watson, pers. comm.). Although the British Museum (N.H.) has no further records concerning the type locality, Ronald S. Wilkinson, a student of the history of entomology, has done some sleuthing to pinpoint the type locality with a high degree of certainty. Given the information that Edward Doubleday probably collected his specimens in the northern U.S. between mid-April and mid-June, Wilkinson reports:
Edward Doubleday and Robert Foster arrived in New York harbor from England on the evening of 24 April 1837. The weather was cold and the season was delayed, so Doubleday amused himself with bird watching, taking only an occasional trip up the Hudson.

On 7 May he left Foster and traveled by riverboat to Albany. After one or two days in Albany he left on 11 May, taking the train to Utica and arriving there in time for dinner that evening. Foster rejoined him at Utica, and the two proceeded immediately to Trenton, about 14 miles north of Utica. They found suitable collecting territory there and decided to remain, making their headquarters about two miles away at Moore's Tavern in Trenton Falls, near the falls of West Canada Creek.

Here Doubleday began to collect moths in earnest. There were no sugar casks available (see Wilkinson, 1866), and, finding the twilight too brief for the usual English method of "nothing," he resorted to two other methods. The first, roaming the surrounding woods with a lantern, was productive, but the mosquitoes were "very annoying" to him. The second, more productive, method of collecting was to put lights in the open windows of the barroom so that moths came in great swarms. Doubleday bottled moths from twilight to ten at night, went to bed, then arose at five or six to spread his captures. This routine went on for months, and he did not stir from Trenton Falls until long after the flight period of *P. assimilans*. Doubleday was still in the U.S. in the spring of 1838, but he was in Florida, far outside the range of *P. assimilans*.

In the absence of data to the contrary, the type locality of *Phragmatobia assimilans* is hereby fixed as the vicinity of Moore's Tavern, Trenton Falls, Oneida County, New York, where Edward Doubleday collected the types in the spring of 1837.

**TYPE SPECIMENS:** Two male syntypes in the British Museum (Natural History), kindly examined for us by Allan Watson.

**DISTRIBUTION** (Fig. 37): In southern parts of the Canadian and northern parts of the Transition Zones, apparently in moist lowlands, from southern British Columbia, central Alberta and central Saskatchewan east through central Ontario to Nova Scotia, south to North Dakota, South Dakota (relict population?), Minnesota, northern Michigan, New York, and New England (except Rhode Island).

Spring collecting with "black" light in suitable localities (marshes or bogs in northern coniferous forest) may extend the known northwestern limit of this species' distribution, and may further result in its discovery in Washington (and perhaps even farther south on the coastal ranges), in the Rockies, and in Wisconsin. It may even occur at high elevations in the Appalachians.

**SPECIMENS EXAMINED (362)**


MASSACHUSETTS. no locality: 16 May, Barnes coll., 1\(^{\circ}\) (USNM).

BARNSTABLE: Barnstable, 17 May 1952, C.P. Kimball, 1\(^{\circ}\) (CNC); 19, 28, 31 May, 1951, 1952, C.P. Kimball, 3\(^{\circ}\) (AMNH). MIDDLESEX: Weston, 22 May 1925, 3 June 1921, 2\(^{\circ}\) (MCZ); C. J. Paine, 1\(^{\circ}\) (Brower); 30 May 1926, 1\(^{\circ}\) (Brower).


EMMET: 5 June 1956, John H. Newman, 1\(^{\circ}\) (MSU).

MACKINAC: T42N, R1W, Sec. 9, 18 May 1964, M.C. Nielsen, 1\(^{\circ}\) (Nielsen).

MONTMORENCY. Atlanta, 30 May 1935, G. W. Rawson, 2\(^{\circ}\) (UMMZ).

OGEMAW: West Branch, 20 & 22 May 1965, Dorothy Frosh, 2\(^{\circ}\) (MSU).

OTSEGO: 26 May 1960, John H. Newman, 1\(^{\circ}\) (MSU).


BRITISH COLUMBIA. Oliver, 5 & 7 May 1923, C. B. Garrett, 2♂ (CNC), 4♂ (AMNH); 14 & 15 May 1953, elev. 1,000 feet, D. F. Hardwick, 5♂ (CNC).


LITERATURE CITED


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Figures 1-9, male genitalia. Figs. 1-3, dorsal view of uncus, anterior to bottom; Figs. 4-6, lateral view of male genitalia from left side, anterior to left (right valva not shown); Figs. 7-9, dorsal view of distal end of aedeagus, anterior to bottom. TOP ROW: *P. lineata*, topotype, 17 Aug. 1964; MIDDLE ROW: *P. assimilans*, Otsego Co., Mich., T29N, R2W, Sec. 18, 22 May 1962; BOTTOM ROW: *P. fuliginosa rubricosa*, Galien, Berrien Co., Mich., 4 Aug. 1965.
Figures 10-19, coremata, valvae, metepisterna, and venation. Figs. 10-12, ventral view of abdomen, coremata shown in situ, anterior to bottom; Figs. 13-15, mesal aspect of right valva, anterior to left, inset a showing cross-sectional outline of valva (hatched) and projection as seen from distal end (right side of figure lifted 90° out of the page) at point indicated by line across valva; Figs. 16-18, plane view of left metepisternum, anterior to top left; Fig. 19, venation of P. fuliginosa rubricosa ♂, no data. TOP ROW: P. lineata, drawn from same male cited on previous plate; MIDDLE ROW: P. assimilans, valva and metepisternum drawn from specimen cited on previous plate, coremata drawn from a second specimen, same locality, 19 May 1962; BOTTOM ROW: P. fuliginosa rubricosa, drawn from specimen cited on previous plate.
Figures 26-33. Figs. 26-29, *P. assimilans*; Figs. 30-33, *P. fuliginosa rubricosa*. Males in left column, females in right. Fig. 26, Lake Kejimukujik, Queens Co., Nova Scotia, 27 May 1958. Fig. 27, Constance Bay, Carleton Co., Ontario, 19 July 1935. Fig. 28, Geraldton, Thunder Bay Dist., Ontario, 11 June 1956. Fig. 29, Hymers, Thunder Bay Dist., Ontario, 1-7 June. Fig. 30, Morenci-Mulberry Rd., Lenawee Co., Mich., 15 July 1961. Fig. 31, Galien, Berrien Co., Mich., 28 July 1965. Fig. 32, Galien, Berrien Co., Mich., 19 July 1965. Fig. 33, Galien, Berrien Co., Mich., 23 July 1965. Photo by Julian P. Donahue.
Figures 34 & 35. Fig. 34, *P. lineata*, holotype male. Fig. 35, *P. lineata*, allotype female. Photo by Julian P. Donahue
Figure 36, distribution of *P. fuliginosa rubricosa*; black circles represent specimens cited in text, triangles represent authentic records in literature or from individuals; the approximate northern limit of forests (tree line) is represented by the bush pattern; scale as for Fig. 37.